

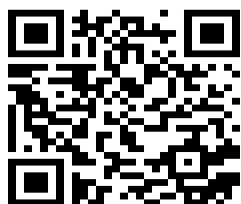


Original Research

Liquid Chromatography-Mass Spectrometry LC-MS Analysis and LC-MS/MS: Application of LC/ESI-MS in In Forensic Sciences, Doping Test, Pharmacokinetics, Bioavailability and Bioequivalence study, Environmental, and Agrochemical and Pesticides Industry

Aqeel Faraj Nahi Farhan Al Bolani¹, Mustafa Abbas Hadi Hamza Al Hashemi², Muntadher Hani Mohsin Abbas Al-Agele³, Laith Sajat Abbas Jabbar Al-Tobi⁴, Shaima Misfer Abdel Aziz Jassam Al Ameri⁵, Abbas Ayyed Radhi AL-Dulaimi⁶

¹⁻⁶Al-Hadi University College, Department of Medical instrumentation Technical Engineering Iraq.



Copyright: ©2024 The Authors. Published by Publisher. This is an open access article under the CC BY-NC-ND license (<https://creativecommons.org/licenses/by-nc-nd/4.0/>).

Abstract:

The technique of LC mass spectrometry is an analytical method that brings together the chromatographic separation of target compounds (or analytes) and the subsequent detection of those compounds based on their mass. In particular, the sensitivity, selectivity, and accuracy of liquid chromatography tandem triple-quadrupole mass spectrometry (LC-MS/MS) have led to the development of procedures that are optimal for detecting nanomolar or even picomolar quantities of a wide variety of analytes. These analytes include drugs, drug and food metabolites, biomarkers of disease progression or drug efficacy, pesticides, food contaminants, markers of ecosystem stability, and natural product extracts. These procedures have been widely utilised because of their unique combination of these characteristics. When it comes to liquid chromatographic separations, the specificity of mass spectrometry detection allows for greater flexibility and efficiency. In the event that quantitative selectivity can be established, the liquid chromatography separation becomes less important for LC/MS/MS than it is for HPLC/UV. This is due to the fact that MS/MS has the ability to detect molecules with differentiable products in a unique manner, even if the molecules co-elute. Simple chromatographic procedures are frequently sufficient for LC/MS/MS, whereas HPLC/UV necessitates complete separation in order to function well. As a consequence of this, the run times that can be achieved using LC/MS/MS can be significantly reduced, and accuracy and precision can be improved. The finest of both worlds is achieved here! Controlling the ionisation process is the most significant obstacle in the identification of multiple sclerosis. When it comes to the MS detector, issues with incomplete ionisation and interference might result in a lack of precision and accuracy. Altering the detection method, utilising a variety of ionisation techniques, utilising sample preparation solvents, utilising LC stationary phases (columns), or LC mobile phases, and utilising chromatographic separations through isocratic or gradient procedures are all potential solutions at your disposal. The time-of-flight MS, the ion capture MS, and other detection methods are also included in the list of other techniques. Electrospray ionisation (ESI), atmospheric pressure chemical ionisation (APCI), photoionization, and other specialised techniques are all

examples of methodological approaches that fall under the category of source ionisation. The precision, accuracy, selectivity, and sensitivity of MS detection are all improved by the use of these one-of-a-kind approaches, which make use of various MS detector setups. Additionally, because it is necessary to minimise the introduction of non-volatile components to the MS source, the choices of LC buffer salts are restricted to volatile buffer systems (typically ammonium cations with formate or acetate anions). These buffer systems may offer limited buffering capacity at typical mobile phase pH levels.

Keywords: LC-MS/MS, LC/ESI-MS, Forensic Sciences, Pharmacokinetics, Environmental, Agrochemical, Pesticides Industry

Introduction:

An increasing demand for precise measurement of microgram and sub-microgram quantities of targets, sometimes in complicated matrices, has been observed in the analytical laboratory. This demand has been observed in a variety of areas, including medicines and food, body fluids, and dirt. Despite the fact that this is not an easy task in and of itself, the possibility of having to analyse hundreds of samples in the shortest amount of time possible while maintaining the quality of the data adds extra hurdles. This powerful instrument has been made available to analytical scientists through the combination of mass spectrometry (MS) and liquid chromatography (LC), which has enabled them to meet these severe standards. Instruments that perform liquid chromatography-mass spectrometry (LC-MS) have become increasingly attractive in many contemporary analytical laboratories as a result of their variety and increased efficiency. Physical separation of target compounds (also known as analytes) is the first step in the LC-MS analytical technique, which is then followed by the detection of the analytes based on their mass. The sensitivity, selectivity, and accuracy of this technique have made it a popular choice for detecting microgram or even nanogram levels of a wide range of analytes, including drug metabolites, pesticides, and food adulterants, as well as natural product extracts. Despite its relatively recent development, this technique has become the method of choice.

A physical separation of the analytes in a liquid sample or a solution of a solid sample can be achieved by the use of liquid chromatography (LC). A few microliters of the sample solution are injected into the mobile phase, which is a stream of a solvent that is moving in a continuous motion. Using an autosampler, it is feasible to accurately inject either as little as 0.1 μL or as much as 100 μL of the sample. The best injection volume is depending on the experimental conditions, however it is possible to inject as little as 0.1 μL or as much as 100 μL .¹ There is a constant pumping of the mobile phase via a column, which is a tube made of stainless steel and is typically filled with silica particles that have been coated with another liquid, which is the stationary phase. As the sample solution-mobile phase mix reaches the column, the components of the sample solution will interact with the stationary phase (which continues to be contained within the column) in a manner that is distinct from one another, depending on the chemical composition or physical qualities of the components. In liquid chromatography (LC) separations, distinct modes have been defined according to the mechanism of interaction between the analyte and the stationary phase. These modes include however:

The separation technique known as partition chromatography is based on the fact that the analytes in the stationary phase have a different degree of solubility and hydrophobicity compared to the mobile phase.

Ion-exchange chromatography is a technique that works by separating analytes according to the ionic charges that they possess.

- Size-exclusion chromatography is a technique that separates analyte molecules by taking use of the differences in the sizes of the molecules for separation.

This technique, known as affinity chromatography, is used to separate analytes according to their capacity to form bonds with the stationary phase.

As the analytes go along the column, they will be separated into their respective categories due to the fact that some of them will interact with the stationary phase more strongly than others. At the beginning of the column's operation, the analytes that have the least amount of interaction with the stationary phase are extracted. The remaining analytes are flushed out in a progressive manner while the mobile phase continues to flow through the column. The analytes that have the strongest interactions emerge last among those that are flushed out. The amount of time that a particular analyte remains in the column is referred to as its retention time (RT), and it is a characteristic of that particular analyte.

The Detection Method of Liquid Chromatography (LC)

The eluent, which is the mobile phase that is flowing out of the column, is then passed through a detector that "responds" to a particular physical or chemical property of the analytes that are contained within it. This characteristic may include the refractive index or the amount of light that is absorbed. It is possible to record this response as a signal or a "peak" whose strength (peak area or peak height) is proportional to the quantity of the component that is present in the sample. RT refers to the moment when the detector "sees" the analyte present in the sample. It is possible to be certain of the identity of a compound that is present in a sample by comparing its RT to the RT of a chemical that is already known. This method of compound identification is not considered to be an accurate method; however, it is helpful in situations where some information about the sample is already known.

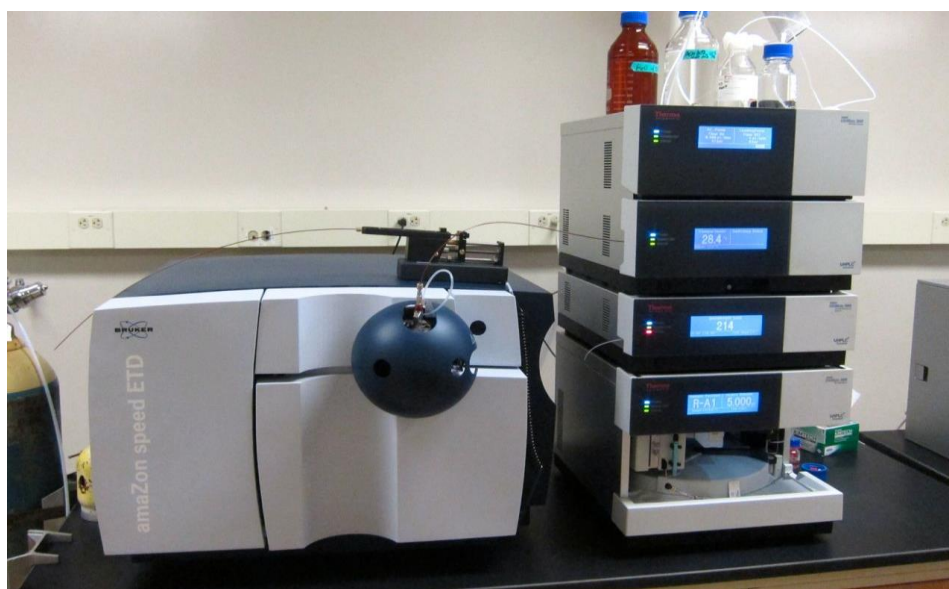


Figure 1. Liquid chromatography–mass spectrometry.

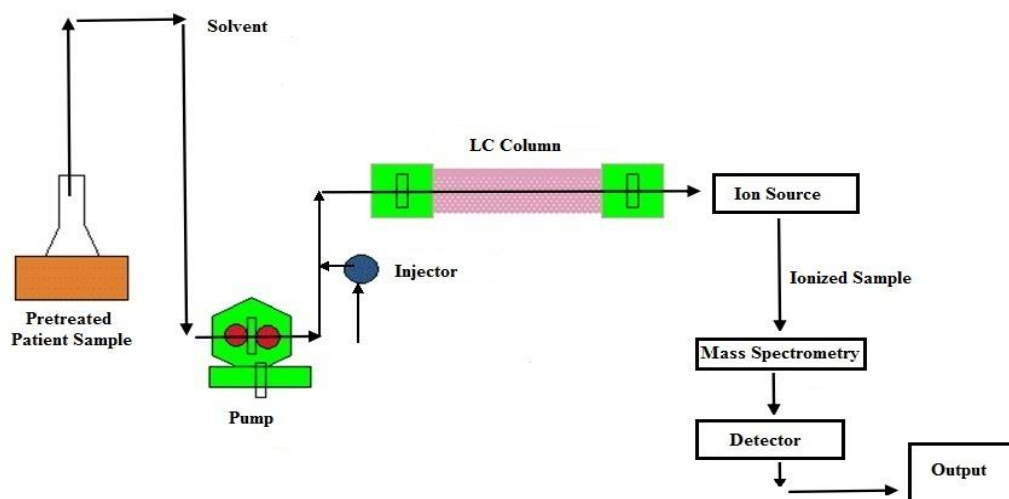


Figure 2. Principle of Liquid Chromatography - Mass Spectrometry (LC-MS)

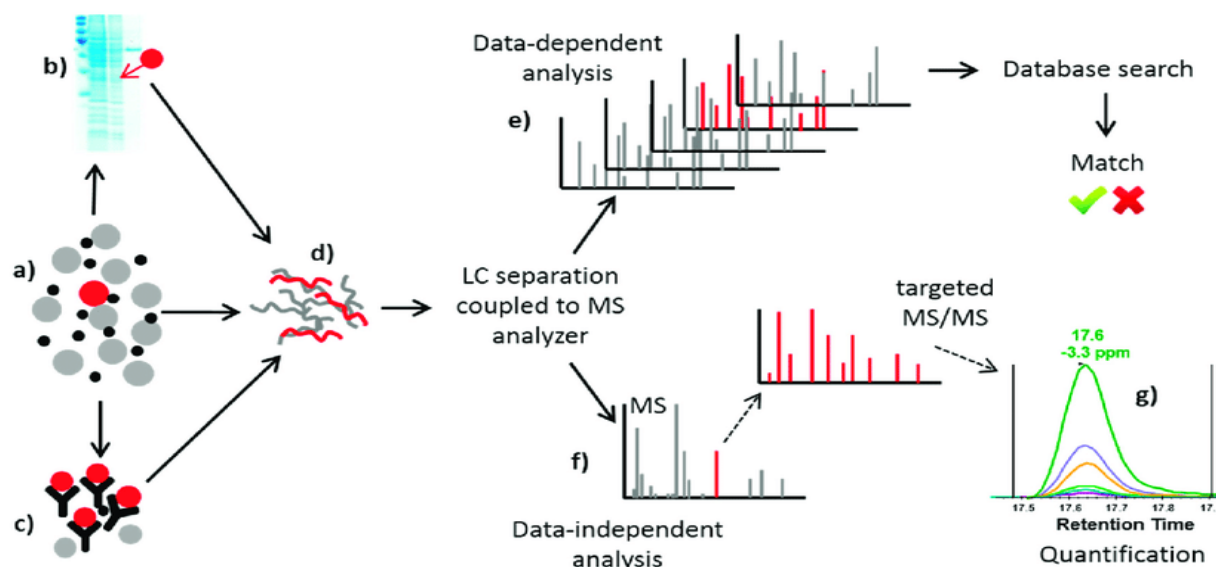


Figure 3. The following are examples of procedures that are based on liquid chromatography-mass spectrometry (LC-MS) for the detection of protein-based toxins: (a) the protein of interest (red) in a complex matrix of other proteins and small chemicals; (b) separation by SDS-PAGE; (c) immunoaffinity purification; (d) enzymatic digest; (e) data-dependent analysis: LC-MS/MS analysis of all of the peptides in the mixture, all of the measured MS/MS spectra are searched against the protein database, and if there is a spectral match, the presence of the agent is confirmed, independent of the amount of the agent that was present. (f) In a targeted (data independent) mass spectrometry/mass spectrometry experiment, peptides of interest are fragmented many times during their elution periods. (g) The extracted ion chromatograms of selected fragment ions are subsequently used for protein quantification (exported from Skyline, unpublished findings).

Utilising MS for the detection of LC

Although a large range of detectors with varying technologies and sensitivities have been combined with liquid chromatography (LC) for the purpose of analysing various types of samples, the mass spectrometer has emerged as a detector that is selective, sensitive, and universal.

The LC eluent that is transporting the separated analytes is not permitted to flow into the mass spectrometer, in contrast to other detectors. The mass spectrometer is often operated in a vacuum, whereas the liquid chromatography system is typically run at ambient pressures. The two systems are coupled together by an interface. The analyte molecules are vaporised and ionised as the column eluent flows into the interface. This process is accompanied by the application of heat, which causes the solvent to evaporate. On account of the fact that the mass spectrometer is only able to identify and measure ions in the gas phase, this is a very important step.

Because the analyte ions are produced at atmospheric pressure in the interface, the process is referred to as atmospheric pressure ionisation (API), and the interface itself is referred to as the API source. Electrospray ionisation (ESI) and atmospheric pressure chemical ionisation (APCI) are the two sources that are utilised in LC-MS analysis the overwhelming majority of the time.

The mass spectrometer is used to attract the analyte ions into it, where they are then subjected to electric forces and/or magnetic fields. Changing the applied fields causes the flight paths of the ions to be adjusted, which in turn ensures that the ions are kept apart from one another according to the mass-to-charge (m/z) values that they possess. It is possible to collect and detect the ions once the separation process has been completed using a number of different mass detectors, the electron-multiplier being the most common of

them. Secondary electrons are emitted when the separated ions come into contact with the surface of the electron-multiplier, which is composed of a dynode. Cascaded through a succession of dynodes, these secondary electrons are multiplied in order to achieve the desired effect. The enhanced current that is produced by the passage of secondary electrons is measured, and the results are connected with the ion concentrations that are present in the mass spectrometer at any given instant in time.

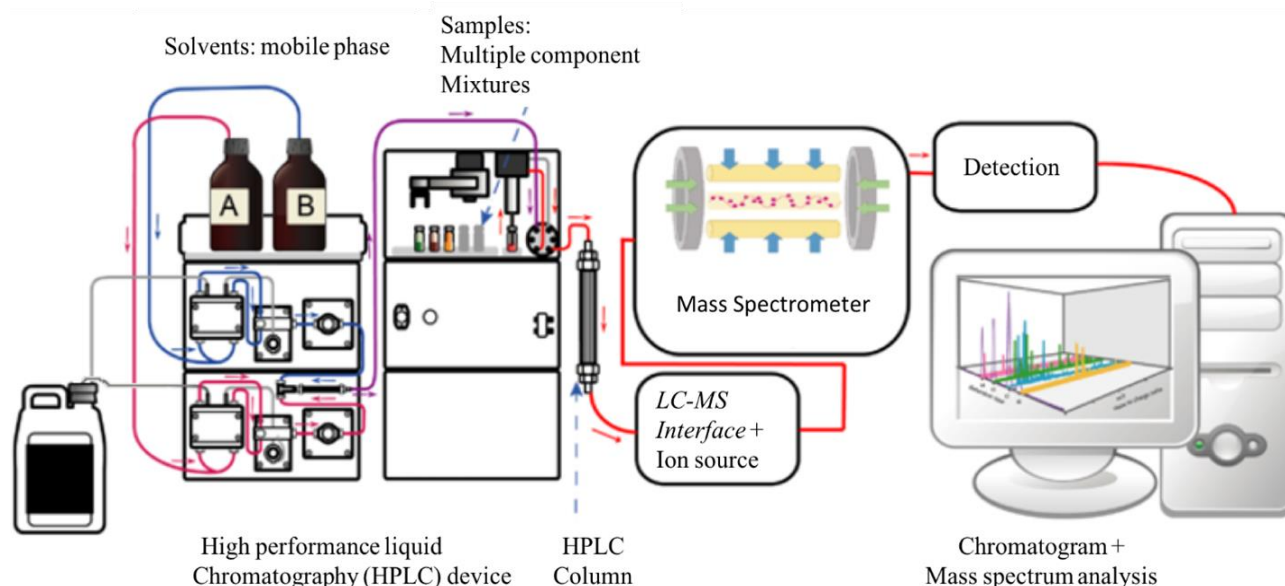


Figure 4. A schematic illustration of an LC-MS setup is provided here. Cwszot, Dagui1929, CasJu, and YassineMrabet are the individuals responsible for this reproduction, which is made available under the Creative Commons CC0 1.0 Universal Public Domain Dedication licence.

.An Analysis of the Data Obtained Through Liquid Chromatography-Mass Spectrometry (LC-MS)

An LC-MS examination of a sample results in the creation of a total ion chromatogram (TIC), which is a plot of the abundances of the ions that were measured during the study. This graph illustrates the peak strengths of the analyte ions in relation to their relative temperatures. To add insult to injury, every single spot on the chromatogram corresponds to a massive spectrum. The ion abundances are depicted in the mass spectrum in comparison to the m/z values that were measured.

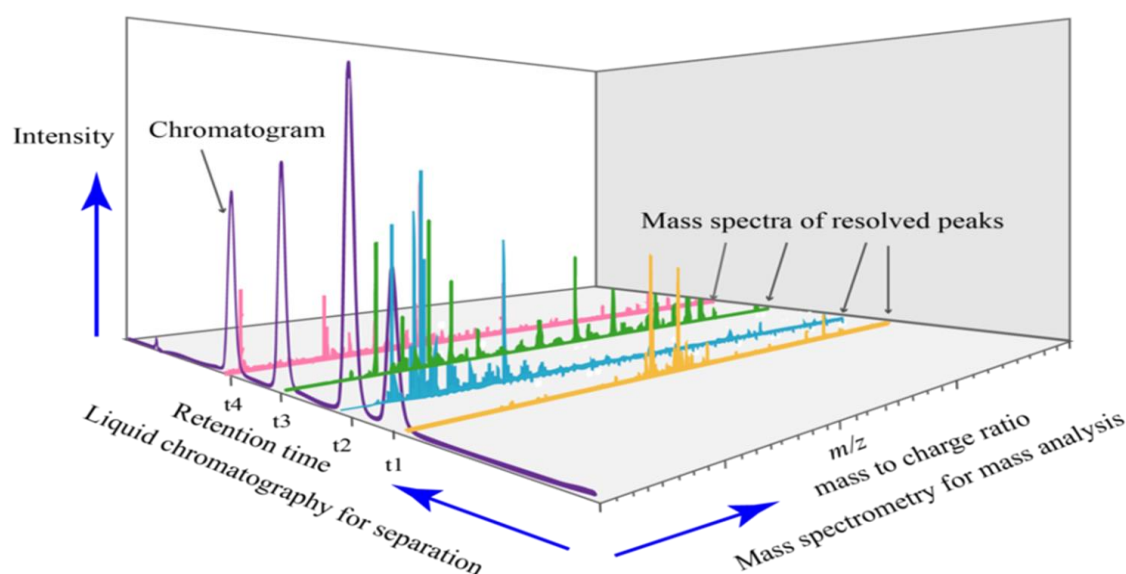


Figure 5. An example of the output plot from the LC-MS laboratory. This image was used with permission from Daniel Norena-Caro and is licenced under the Creative Commons CC0 1.0 Universal Public Domain Dedication..

The mass spectrum of a compound not only provides information about the mass of the parent molecule (based on the m/z value of its ion), but it also helps to understand the structure of the compound by analysing the relative abundances of isotopic mass peaks. This information is obtained from the mass spectrum of the compound. The area of the peak of the analyte is utilised for the purpose of quantifying it. Scan and selective ion monitoring (SIM) are the two modes of operation that are available for the mass spectrometer at any given time. When it is in the scan mode, it is configured to identify all of the ions within a given time period, ranging from low m/z values to high m/z values altogether. In situations where there is no information available regarding the ions that are present in a sample or when you are analysing samples that are unknown, this mode is utilised. The mass spectrometer is configured to measure particular m/z values when it is operating in SIM mode. This is the mode of operation that is recommended for precise quantification of substances that are already known to be present in a sample.

The procedure of combining liquid chromatography with tandem mass spectrometry, often known as LC-MS/MS

By combining two mass analyzers that are run in series, it is possible to obtain additional improvements in the identification of samples and the precision of their quantification. Among the tandem mass spectrometers, the ones that are utilised the most frequently are the triple quadrupole mass spectrometers (QQQ or TQMS) and the quadrupole time-of-flight (QTOF) models. With these arrangements, there are multiple options available for conducting sample analysis.

There are two quadrupole mass analyzers (Q1 and Q3) that make up the TQMS. These analyzers are separated from one another by a collision cell (q/Q2). The collision cell is used to fragment the precursor ions that were isolated in Q1 by subjecting them to high-energy collisions with a neutral gas, such as argon, helium, or nitrogen. While Q1 and Q3 are used as mass analyzers to scan over a mass range or to monitor an ion with a specified m/z value, the collision cell is used to fragment the precursor ions. It is possible to run a Total Quality Management System (TQMS) in four distinct modes(5), which are as follows:

- Precursor ion scan: the first quadrupole (Q1) is scanned over a mass range in order to choose the precursor of a particular product ion (m/z value), which is then monitored in the third and final quadrupole (Q3).
- Product ion scan – Q1 is configured to send just the pre-defined precursor (m/z) to the collision cell, whilst Q3 is scanned over a mass range in order to identify the fragments that were obtained under the experimental conditions.

Neutral loss (NL) is a method that involves scanning both Q1 and Q3 in order to identify all of the precursors that are responsible for the production of the products. This is accomplished by removing the identical neutral (uncharged) species from each of the precursors. There is a difference between the scan range of Q3 and the NL value.

- Selective reaction monitoring (SRM) - both Q1 and Q3 are configured to monitor particular m/z values for the ions that are associated with the precursor and the product. When it comes to chemical quantification, this approach is the one that is recommended because of its specificity and sensitivity. In addition to monitoring multiple precursor-to-product transitions of the same analytes, TQMS can also be used to monitor transitions between different analytes.

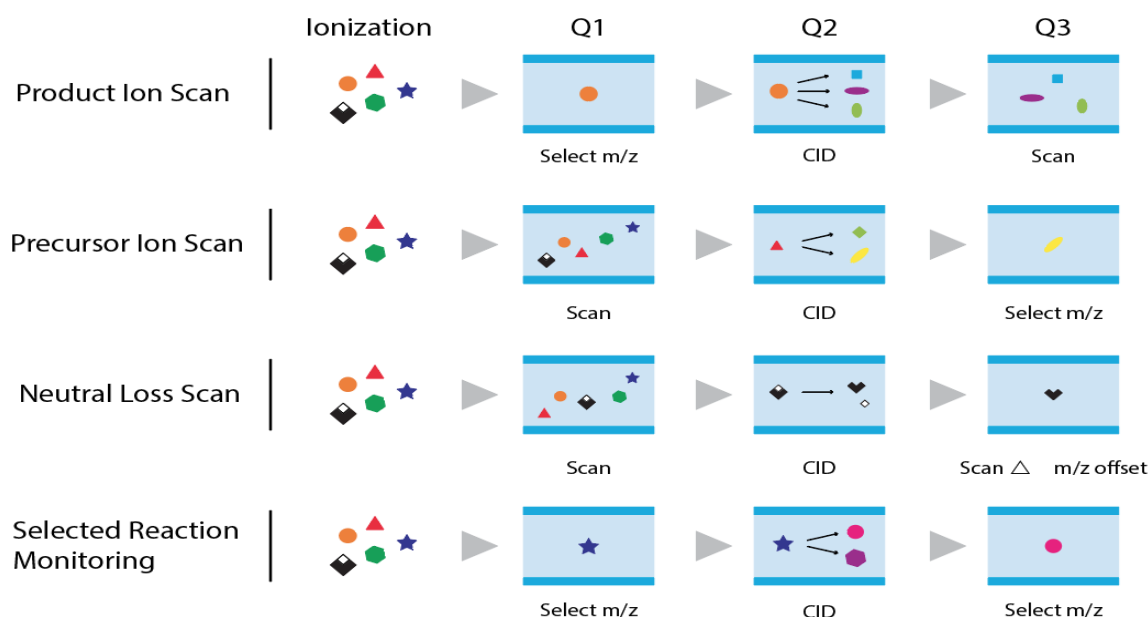


Figure 6. Modes of operation of TQMS.

The fragmentation is determined by the structure of the molecule as well as the conditions of the experiment, which include the pressure of the gas and the energy of the collision. On account of this, the fragmentation pattern, in conjunction with the compound RT and the precise mass value of the compound, is utilised for the purpose of identification under a certain reaction scenario. In addition, the monitoring of particular fragment ions helps to improve the sensitivity of detection, which in turn makes it possible to quantify smaller quantities of the chemicals that are being targeted. The quadrupole mass analyzer and the time-of-flight mass analyzer that are contained within the QTOF mass spectrometer are separated from one another by a collision cell. With the quadrupole, it is possible to either transmit the ions or isolate a particular precursor ion, which is subsequently fragmented in the collision cell. Both of these applications are possible. After being pulsed into the TOF analyzer by a modulator, a small portion of the ions are then accelerated into the high-vacuum field-free area by providing high voltage. This process is repeated until the entire ions are accelerated. Ions that have m/z values that are different from one another go down the flight tube at different speeds and continue to be separated from one another. High mass resolutions are provided by TOF mass analyzers, which also have the capability to scan over vast mass ranges in a short amount of time.

Quantitative analysis using liquid chromatography and mass spectrometry (LC-MS)

A wide variety of matrices have been subjected to LC-MS analysis, which has been utilised extensively for the investigation of both small molecules and large protein molecules. Applications of this technology include, but are not limited to the following:

- the detection of twelve model chemicals that represent certain kinds of doping agents, such as anabolic agents and simulants, in exhaled breath;
- the measurement of drug metabolites in bodily fluids; and
- (4) the quantification of genotoxic contaminants discovered in active pharmaceutical ingredients.

The detection of adulterants in food materials and dietary supplements, the determination of alkylphenol ethoxylates (APEOs) in tannery sediments, the quantification of personal care products in swimming pool and river water samples, the quantification of nucleotides and their derivatives in bacterial cells, and the quantification of the proteome are all applications of this technique.

a fast technique for the identification of SARS-CoV-211 as a potential vaccine

In addition, the method has been utilised in the examination of drinking water, petrochemicals, soil, biopharmaceuticals, food, and wine, as well as in the detection of per- and polyfluoroalkyl substances (PFAS) and pesticide residues.

Considerations Regarding the Benefits and Drawbacks of Liquid Chromatography-Mass Spectrometry (LC-MS)

Aspects of Strength

LC-MS is an analytical technique that can be utilised for the examination of thermolabile molecules, as well as polar and non-polar substances. Low molecular mass analytes with m/z values less than 1000 Da and very high molecular mass proteins with m/z values greater than 100,000 Da are examples of the types of chemicals that can be found in this category. The "soft ionisation" of the compounds primarily results in the production of the molecular ion as well as the isotopic peaks, both of which are beneficial in the process of determining the precise mass and putative formula of the analyte by the instruments. When the fragmentation spectra are taken into consideration, it is feasible to gain a better understanding of the structure of the analyte.

Even though the LC-MS technique requires only a few milligrammes of a pure substance in order to do an analysis, it is possible that as little as one milligramme is needed. It is feasible to reach limits of detection in the range of ng/mL or even pg/mL by operating the MS in SIM or SRM mode. This is possible so that the MS can be used. Even for analytes that are present in complex matrices, selectivity can be accomplished through the use of SRM mode since it exclusively monitors particular ions.

The constraints

The ownership, operation, and maintenance costs of LC-MS devices are quite high. For the purpose of operating the instruments and analysing the data, expertise is necessary. Comparatively speaking to other analytical methods, the sample throughput is considered to be modest. Due to the fact that the spectra obtained are dependent on the experimental conditions, including the type of equipment, the scope for compound identification through comparison to the reference spectra is restricted. Due to the fact that a mass spectrometer is a destructive detector, it is imperative that caution be exercised when working with materials that may not be easily accessible or that cannot be obtained in significant quantities. The analysis of unstable or reactive samples with LC-MS can be difficult because it is a technique that is performed in the laboratory rather than as an in-field approach.

Therefore, solid samples must either be dissolved in an appropriate solvent or the analyte must be extracted from the sample in order to be injected into the column. This is because the column can only accommodate liquid samples. In order to successfully extract target analytes from complex materials including blood plasma, food, and soil, it is necessary to first prepare the samples using procedures such as liquid-liquid extraction (LLE) or solid-phase extraction (SPE).¹² Not only does this serve to improve the sensitivity of the analysis, but it also helps to limit the potential for contamination inside the system, which will be explored in the following section.

Instances of Liquid Chromatography-Mass Spectrometry (LC-MS) that are frequently seen

In spite of the fact that LC-MS offers a number of benefits for trace analysis in complicated matrices, there are a number of precautions that need to be followed in order to overcome the following challenging situations while utilising this method.

A contamination of The sensitivity, selectivity, reproducibility, and resolution of an analysis are all affected by contaminants. These contaminants include metal ions, phthalates, polyethylene glycol (PEG), slip agents, water, and particles that enter the system from a variety of sources. These sources include: reagents and

solvents; water used for buffer preparation; chemicals leaching from glassware; microcentrifuge tubes; inlet filters; solvent lines.

- components of the instrument, such as pump seals
- gases that are utilised for the desolvation of the eluent at the source and in the collision cell
- the sample itself

It is possible for the contaminants to cause interference with the analysis in the following ways: by suppressing or boosting the ionisation of analyte(s) in the source; by forming adducts with the analytes; by obscuring the analyte peaks and/or appearing as ghost peaks in the chromatogram; and by making the baseline noisy.

- causing the system and the column to become clogged, which necessitates regular maintenance and the replacement of components

In order to reduce the risk of contamination, it is recommended that high-purity solvents, water, and reagents be utilised in the process of preparing mobile phases.

- Mobile phases that have been freshly prepared must be utilised in order to reduce the likelihood of microbiological contamination of aqueous mobile phase and polymerization of acetonitrile (ACN).

It is recommended that you refrain from using soap or detergent to clean glassware because these substances can be difficult to remove and can create interference during the analysis process.

- It is required to make use of gases with a high purity, such as nitrogen gas, which is often utilised and has a purity of more than 95%.
- Nitrogen generators need to be carefully maintained, and the gas cylinder has to be replaced when the pressure drops below the appropriate level.
- Analytes need to be extracted from the sample matrix, and chromatographic parameters need to be optimised in order to improve the resolution of analyte peaks in comparison to interfering peaks.

Those Matrix Effects

It is possible for other constituents of the sample to either inhibit or enhance the ionisation of the analyte in the source when conducting an analysis of biological samples. It is recommended that the analytes of interest be separated from the matrix in order to reduce the influence of the matrix. As a result, the preparation of samples is an essential prerequisite for LC-MS analysis. This may make it more difficult to extract only the analyte(s) from the matrix, despite the fact that it lowers the effects of the matrix. The chromatographic settings can also be optimised in order to prevent the co-elution of chemicals that interfere with the separation process. Another method that helps to account for matrix effects is the preparation of standard solutions in an analyte-free matrix, which is known as matrix matching. To compensate for the matrix effects, known amounts of isotopically labelled internal standards are utilised. These standards undergo ionisation suppression or enhancement in a manner that is comparable to the matrix effects.

Continued use

Because of sample carryover, it is possible for analyte peaks to be observed in blank injections that are performed after a sample with a high concentration. It is necessary to solve this issue by implementing cleaning processes, which include multiple blank injections, needle washing, and column conditioning, in order to guarantee that the sensitivity of the analysis is preserved.

Loss of Samples

It is possible for non-specific binding to laboratory consumables, such as the inner surface of microcentrifuge tubes, to result in the loss of analytes. Some examples of these analytes include proteins and

DNA. There is a correlation between the adsorption of analytes and the accuracy and precision of the assay. It is possible to reduce the amount of analyte that is lost by utilising containers that have a low surface adherence. Adding blocking agents that reduce the amount of interaction that the analyte has with the interior surfaces of the containers is yet another method that can be utilised.

The choosing of the mobile phase buffer

Only volatile buffers, such as ammonium formate or ammonium acetate, which do not precipitate in the source can be used for the production of the mobile phases. This is because the column eluent needs to be removed before the MS analysis can be performed.

To keep up with

It is necessary to perform routine maintenance on the mass spectrometer in accordance with a predetermined schedule in order to guarantee the instrument's precision, reproducibility, and trouble-free operation, as well as to reduce the amount of unscheduled downtime that occurs. With the help of LC-MS, sensitive quantification of drug metabolites is possible.

In the process of developing new drugs, one of the most important considerations is figuring out what happens to a medicine once it has been absorbed by the body. It is impossible to evaluate the efficacy and safety of a medicine without first examining its metabolic destiny and gaining an understanding of the pharmacological responses that come from this. The detection and quantification of metabolites, on the other hand, is not a straightforward operation. This is particularly the case because of the uncommon presence of metabolites in biological samples and the intricate nature of the matrices that are involved. From this point of view, liquid chromatography coupled with mass spectrometry, also known as LC-MS, has emerged as one of the most potent analytical methods for the screening and identification of drug metabolites.¹. Recent developments in the field of liquid chromatography-mass spectrometry (LC-MS) have enabled researchers to conduct more in-depth analyses of drug metabolic pathways and drug-drug interactions, as well as to enhance therapeutic options for a wide range of difficult disorders.

In comparison to Liquid Chromatography-Mass Spectrometry (LC-MS/MS), High-Performance Liquid Chromatography (HPLC) is a technique that

employing liquid chromatography-tandem mass spectrometry (LC-MS/MS) techniques, Dr. Mohd Yusmaide Aziz, a researcher at the Advanced Medical and Dental Institute in Universiti Sains Malaysia, has spent years employing these techniques to investigate the metabolites of anti-malarial drugs like piperazine. Through his explanation, he adds that when it comes to quantifying metabolites, pharmaceutical scientists frequently favour LC-MS/MS over other analytical techniques due to the fact that it is both applicable and sensitive.

According to Aziz, "high performance liquid chromatography (HPLC) was commonly used for drug quantitation a decade ago because of its capabilities." In order to identify compounds, either ultraviolet (UV), fluorescence, or electrochemical detection was utilised. However, in today's world, researchers in the pharmaceutical industry have access to more advanced technologies such as mass spectroscopy, which possesses increased sensitivity, resolving power, and a broad dynamic range. When working with restricted biological samples, sensitivity is a crucial factor. According to Aziz, the LC-MS/MS technologies that he utilises provide high enough selectivity and sensitivity to analyse the parent drug together with its metabolites. This is especially true when working with limited biological samples. According to him, "During the LC-MS/MS operation, a compound will be ionised and fragmented to specific product ions of the compound, and this is considered to be a fingerprint." The technique enables molecules to be detected at the parts per trillion (PPT) level, which provides a means for Aziz's team to measure the concentration of

piperaquine and its metabolites in human plasma. These metabolites are even lower than the concentration of the parent drug. "In addition, LC-MS/MS has a very high level of sensitivity to detect trace amounts of the compound," the researcher said.

Assessing the Metabolites of Anti-Malarial Therapeutics

When piperaquine enters the body, it is metabolised in the gut and the liver to compounds that have half-lives that are equal to or longer than those of piperaquine. These compounds can sometimes last for three to four weeks. "Our interest in quantifying piperaquine metabolites stemmed from the fact that none of these compounds had previously been assayed in systemic circulation," explains Aziz. Piperaquine is an anti-malarial drug that is commonly used in combination with dihydroartemisinin due to its long biological half-life. "Determining the concentrations of piperaquine in human plasma can help us to further study the effects of metabolites in systemic circulation and their potential for drug-drug interactions even after the treatment has ended." Aziz and his collaborators at Sahlgrenska Academy, University of Gothenburg, Sweden, developed a sensitive and quantitative assay for piperaquine and two of its metabolites. The assay was developed using liquid chromatography-mass spectrometry. The technique involved separating analytes from plasma samples on a C18 column and detecting them on a tandem mass spectrometer with an ESI source operated in the positive ion mode with deuterated piperaquine as an internal standard.³ Detection and quantitation was based on the mass-to-charge ratios (m/z) of precursor-product ion pairs, which are specific to piperaquine and its metabolites. "Before running samples, we made sure to validate the method based on FDA guidelines for bioanalytical methods," says Aziz. When constructing the approach, it is imperative that the criteria outlined in the guidelines be adhered to in a careful and meticulous manner. Some of the parameters that are considered are accuracy, precision, stability, recovery, and matrix effects.

A Quantitative Analysis of HIV Treatment Metabolites

A combination of two nucleoside reverse transcriptase inhibitors (NRTIs) is the first line of treatment that is recommended for individuals who are suffering from acquired human deficiency syndrome (AIDS), which is caused by human immunodeficiency virus-type 1 (HIV-1).⁴ In essence, these molecules have the ability to prevent HIV from producing copies of itself when it enters a healthy human cell. This mechanism is accomplished by NRTIs through the inhibition of an enzyme known as reverse transcriptase; however, before this can occur, the molecules must first be metabolised to their active form. According to Dr. Yazen Alnouti, an associate professor in the Department of Pharmaceutical Sciences in the College of Pharmacy at the University of Nebraska Medical Centre (UNMC), non-retroviral anti-inflammatory medicines (NRTIs) are also known as prodrugs. Before they are able to inhibit HIV reverse transcriptase, they must first undergo intracellular phosphorylation to become active triphosphate (TP) nucleotide metabolites. It is challenging to monitor these pharmacologically active metabolites inside of cells because of their hydrophilicity, instability, and low concentrations in blood and tissues. Due to the hydrophilicity of TP metabolites, traditional reverse phase chromatography is not an effective method for separation. As a result, alternative methods are utilised, which can be classified as direct and indirect LC-MS methods.⁵ The direct method is based on the direct quantification of the nucleotide metabolites under conditions that are not associated with reverse phase liquid chromatography. On the other hand, the indirect approaches entail measuring the parent nucleosides by the dephosphorylation of the metabolites when the circumstances are in reverse phase liquid chromatography. The team led by Alnouti explored both direct and indirect methods for quantifying the non-retroviral antiretroviral drugs (NRTIs) lamivudine (3TC) and abacavir (ABC), as well as their metabolites. According to Alnouti, "the majority of the existing methods that measure TP metabolite concentrations in peripheral blood mononuclear cells (PBMCs) require sourcing these cells from large volumes of human blood, which range from six to eighteen hundred millilitres." We had the objective of

developing and validating a sensitive and selective LC-MS/MS approach that is capable of quantifying TP metabolites in peripheral blood mononuclear cells (PBMCs) that originate from significantly smaller blood volumes (~0.5 ml).

Method for the Sensitive Measurement of NRTI Metabolites Utilising LC-MS/MS

Researchers working under the direction of Alnouti at the University of New Mexico were able to quantify metabolites from two different NRTI medications in mouse blood at the same time. In addition to this, they measured the concentration of these metabolites in mouse immune cells that were taken from the spleen, liver, and lymph nodes. The lead author of the study, Dr. Nagsen Gautam, now an assistant professor at UNMC, clarified that their technique involved isolating TP metabolites using anion exchange chromatography, converting them to the parent nucleosides via dephosphorylation, and analysing these parent nucleosides as surrogates for their TP metabolites using a sensitive LC-MS/MS method.⁶ He further explained that for targeted quantification of small metabolite molecules, most triple-quadrupole LC-MS/MS systems in multiple reaction monitoring mode (MRM) will grant high sensitivity and selectivity. The lower limits of quantification for the 3TC metabolite were found to be 10 pg/ml, while the lower limits of quantification for the ABC metabolite were found to be 4 pg/ml.⁶ According to Gautam, these results suggest that the method that his team developed is 12.5 to 50 times more sensitive than the direct and indirect LC-MS/MS methods that were previously used to measure 3TC and ABC triphosphate metabolites. "In addition to optimising sensitivity, we also optimised conditions to assure the stability of TP samples and standards during sample collection, preparation, LC-MS/MS analysis, and storage. This was done simultaneously with the optimisation of sensitivity. In general, these aspects are not taken into consideration by other approaches," he went on to say. The specific conditions are detailed in a paper that was just recently published in the Journal of Pharmaceutical and Biomedical Analysis by Gautam.⁶ The availability of a precise method to monitor the levels of pharmacologically active metabolites in limited blood and tissue samples will assist in the acceleration of the preclinical and clinical development of antiretroviral therapy that is more effective for patients who are afflicted with HIV/AIDS. Alnouti's team is currently concentrating its efforts on locating TP metabolites in inaccessible viral reservoirs throughout the body, armed with the LC-MS technique that they have perfectly optimised. Alnouti argues that the release of the virus from hidden viral reservoirs, primarily lymph nodes, is the cause of HIV resistance and recurrence after years have passed when the infection has been present. It is possible to support the optimisation of new therapeutics by monitoring active intracellular TPs in these hidden reservoirs. These therapies have the ability to attack the virus in these concealed places and entirely destroy the infection.

Within the realm of forensic sciences, the application of LC/ESI-MS

The LC-MS technique is utilised for the purpose of determining toxicity, as well as for drug analysis and trace analysis. The LC-MS technique can be used to determine the presence of toxins in a variety of materials by using a tiny amount of sample. It is possible to determine whether or not food or beverages contain any hazardous metabolites by utilising LC-MS. For instance, the identification of detergent that was added to orange juice can be determined by doing an analysis on the sample of both the juice and the detergent. In this case, alkyl diphenylether sulphonic acid, which is the usual surfactant, is utilised. Samples of juice and detergent are subjected to the same chromatographic conditions for analysis. The reference spectra of the standard surfactant, which is alkyl diphenyl ethersulfonic acid, are identical to the mass chromatograms and mass spectra that were obtained from the juice and detergent samples [25,26].

Analysis of Doping Using LC-MS as an Application

In order to identify the presence of the 4-Methyl-2-hexanamine doping agent in urine, the LC/ESI-MS coupled with the positive mode can be utilised. While the urine samples are being examined, an internal

standard consisting of tuaminoheptane is also being added. The primary amine 4-methyl-2-hexaneamine, which is an analogue that is found in nutritional supplements, is thought to be the unknown molecule. Four-methyl-2-hexaneamine is the standard that is utilised, and it displays two peaks that are not resolved at RT 3.43 minutes and 3.78 minutes. which are identical to those of the chemical that is unknown [26]. It was possible to identify 4-methyl-2-hexaneamine using the single reaction monitoring (m/z 116-57), which was a specialised method.

Application: Within the field of pharmacokinetics:

A technique known as LC-MS is utilised in the investigation of drug absorption, metabolism, and excretion. Bio analytical procedures are utilised for the purpose of elucidating the quantitative and structural characteristics of pharmaceuticals and their metabolites in biological samples such as plasma, urine, saliva, serum, and other similar substances [25].

When it comes to the study of bioavailability and bioequivalence's applications:

pharmacodynamics, clinical trials, and in-vitro dissolution tests are examples of comparative bioequivalence investigations [27,28]. These studies involve the quantitative assessment of medicines or metabolites in biological matrix.

In the process of determining the molecular weights, applications include:

LC-MS is utilised for the purpose of determining the molecular weights of substances that are either known or unknown. It gives information about the molecular weight, structure, identification, and quantity of the components in the sample. When it comes to determining the molecular weights of proteins, nucleic acids, polymers, and peptides, LC-MS is something that is utilised.

For the purpose of determining the assay of drugs and intermediates, applications include:

In the pharmaceutical sector, liquid chromatography-mass spectrometry (LC-MS) is utilised for the purpose of determining the assay of drug substances, drug products, intermediates, and corresponding chemicals [3].

The agrochemical and pesticide industries are two examples of applications.

Additionally, it is utilised in the process of determining the various components that are included in pesticides and fertilisers [25].

Environmental Applications: Applications in the Environment

The detection of phenyl urea herbicides and the detection of low levels of carbaryl in food are both possible with the help of LC-MS. A. L. Perrenoud " developed LC-MS approach for the detection of 4-methyl-2-hexaneamine, a doping substance, from urine. The method uses LC-MS with ESI in positive mode. The single reaction monitoring (m/z 116-57) demonstrates that the analyte was specifically detected for the presence of 4-methyl2-hexaneamine. The analyte was separated using a gradient mobile phase on some reverse phase C8 column. A mass spectrometry technique known as atmospheric pressure photoionization (APPI) of guanine was developed by Allegrand J. utilising synchrotron VUV light that could be tuned. Within this APPI source that is connected with a VUV photon source that may be tuned. The ionisation of guanine was brought about by chemical processes, and the energy of the photons played a role in this process. The LC-MS method was created by Pascual-Teresa Standard for the purpose of analysing anthocyanins derived from purple maize cobs. A technique known as LC-MS is used to separate and identify each of the nine distinct forms of anthocyanins. Using fragmentation patterns (MS spectra), this liquid chromatography (LC) experiment, which was linked with diode array spectrometry and mass spectrometry, was able to determine the anthocyanins components that were present in purple maize cobs. For the purpose

of determining the total amount of resveratrol present in grape juice, cranberry juice, and wine, Wang Y devised an LC-MS method. Reverse phase high-performance liquid chromatography (HPLC) with positive ion atmospheric pressure chemical ionisation (APCI) mass spectrometric detection was utilised in order to evaluate the samples. Grape juice, cranberry juice, and wine were all shown to contain resveratrol, with concentrations ranging from 1.56 nmol/g, 1.07 nmol/g, and 8.63 to 24.84 $\mu\text{mol/L}$, respectively. A description of the most recent advancements in LC-MS for pharmaceutical analysis is provided by Chang-Kee L. Electrospray, atmospheric chemical ionisation, and photo ionisation are some of the techniques that are addressed in this article, along with their respective interfaces. An overview of the applications of liquid chromatography-mass spectrometry (LC-MS) in drug development, in vitro and in vivo drug metabolism, identification, and characterisation of impurities in pharmaceutical analysis has been provided. Within the field of forensic toxicology, Nishikawa M. described the utilisation of LC-MS for the measurement of surfactants. The analysis of anionic, cationic, and non-ionic surfactants is carried out in both the negative and positive modes. This means that anionic surfactants and positive surfactants are detected as M^- ions in the negative mode and M^+ ions in the positive mode. On the other hand, non-ionic surfactants are detected as $[\text{M}+\text{H}]^+$ ions or $[\text{M}+\text{NH}_4]^+$ ions in the positive mode. It was found that the recovery range for anionic, cationic, and non-ionic surfactants was between 65.8% and 124%. Through the utilisation of LC-MS, Hernando MD was able to ascertain the minute amount of pharmaceutical residues present in both natural and treated water. The samples, which include wastewaters from rivers and taps, as well as wastewaters from influent and effluent, are examined. Ibuprofen, Ketoprofen, and Diclofenac are some examples of medicines that can be detected at the trace level by the utilisation of solid-phase extraction (SPE) in conjunction with liquid chromatography tandem mass spectrometry. The quantitation limit and the detection limit of the technique were both between 7.5 and 75 ng/L. By utilising LC-ESI-MS, Souverain S was able to establish a method for determining the formation of protein precipitation in plasma for the purpose of conducting drug cocktail analysis. Acetonitrile (ACN), perchloric acid (PA), and trichloroacetic acid (TCA) are the three acids that are utilised in the process of protein precipitation (PP). For the purpose of conducting simultaneous analysis of six different substances in a time span of less than six minutes, the LC-ESI-MS technology was created. Depending on the protein precipitation techniques that are successful in removing protein from human plasma and compatibility with LC-ESI-MS, ACN is employed as a PP technique. This technique ensures a recovery of more than 80% and a CV of up to 6%. The LC-ESI-MS approach was developed by Bogusz MJ for the purpose of identifying synthetic adulterants that are present in herbal treatments. Drugs are extracted using methanol, and the separation process is carried out using liquid chromatography with a gradient mobile phase consisting of acetonitrile and a buffer containing 10 mM ammonium formate at a pH of 3.0. In each injection of material, the limit of detection that has been recorded is between 5 pg and 1 ng. The percentage of recovered medications that have been spiked ranges from 63 to 100 percent. Roach AGS provides a description of the application of the LC-MS/MS technology for the determination of acrylamide for use in food. As a standard for internal use, acrylamide is utilised. 10 parts per billion ($\mu\text{g/kg}$) is the limit of quantification for [13 C3]. Numerous types of dishes can be prepared using this method. The acrylamide content in foods can be reduced thanks to the fact that the analyte levels in different types of foods might vary. The LC-MS method for simultaneous estimation of amlodipine and valsartan in human plasma was developed and validated by Jangala H.; this approach was used to do the estimation. Using an isocratic technique with a mobile phase consisting of acetonitrile and a solution of 5mM ammonium formate (80:20 volume/volume), the chromatographic separation was accomplished at a flow rate of 0.8 millilitres per minute. Under the conditions of multiple reactions monitoring (MRM), the quantification was carried out using enhanced spectroscopy (ESI) in positive mode. When it comes to amlodipine, the assay yielded a range of 0.302-20.725ng/mL, whereas the range for valsartan was 6.062-18060.792 ng/mL.

Conclusion:

LC-MS, which stands for liquid chromatography-mass spectrometry, is an extremely effective analytical method that possesses both a high level of sensitivity and of specificity. Liquid chromatography (LC) and mass spectrometry (MS) are the two components that make up the LC-MS technique. The separation of components can be accomplished by the use of liquid chromatography (LC), and the sample eluents from LC are then transferred into mass spectrometry (MS), which allows for the detection, identification, and measurement of the masses of components even when other components are present. In the pharmaceutical industry, LC-MS is utilised for the purpose of determining pharmaceutical drug ingredients, intermediates, and related molecules for both quantitative and qualitative purposes. In-vitro dissolution, bio-equivalence, bioavailability, and metabolite investigations are the areas in which LC-MS is utilised the most for significant purposes. LC-MS is also utilised in the fields of basic research, agrochemical, and forensic laboratories, as well as the food industry. It is a hyphenated approach that is employed in conjunction with the separation power of high-performance liquid chromatography (HPLC) and the detection power of mass spectrometry (MS). Applications in the fields of forensics, environmental science, and the pharmaceutical, chemical, food, and agrochemical industries are among its most common uses. Liquid chromatography-mass spectrometry (LC-MS) is utilised for the qualitative and quantitative analysis of biological materials and pharmacological compounds. In the field of medication research and quality control, it is also frequently utilised.

References:

1. Dolan JW, Snyder LR. (1989) Injectors and autosamplers. In: Troubleshooting LC Systems. Humana Press, Totowa, NJ. doi:10.1007/978-1-59259-640-9_10
2. Medhe S. Mass spectrometry: Detectors review. Chem. Biomol. Eng. (2018); 3(4):51–58. doi:10.11648/j.cbe.20180304.11
3. Markgraf DF, Al-Hasani H, Lehr S. Lipidomics—Reshaping the analysis and perception of Type 2 diabetes. Int. J. Mol. Sci. 2016;17(11):1841. doi: 10.3390/ijms17111841
4. Chidella K, Dasari VB, and Anireddy J. Simultaneous and trace level quantification of five potential genotoxic impurities in ranolazine active pharmaceutical ingredient using LC-MS/MS. Am. J. Analyt. Chem, 2021;12:1–14.
5. Thevis M, Krug O, Geyer H, Schänzer W. Expanding analytical options in sports drug testing: Mass spectrometric detection of prohibited substances in exhaled breath. Rapid Commun. Mass Spectrom. 2017;31(15):1290–1296.
6. Frank N, Bessaire T, Tarres A, Goyon A, Thierry Delatour. Development of a quantitative multi-compound method for the detection of 14 nitrogen-rich adulterants by LC-MS/MS in food materials, Food Addit. Contam: Part A. 2017;34(11):1842–1852.
7. Dincel D, Olgan H, Canbaloglu Z et al. Determination of dihydrocapsaicin adulteration in dietary supplements using LC-MS/MS. J. Chem. Metrol. 2020; 14(1):77–82. doi: 10.25135/jcm.36.20.01.1532
8. Mahalakshmi R, Pugazhendhi A, Brindhadevi K, Ramesh N. Analysis of Alkylphenol ethoxylates (APEOs) from tannery sediments using LC-MS and their environmental risks. Process Biochem. 2020;97:37–42, ISSN 1359–5113, doi: 10.1016/j.procbio.2020.06.015
9. Kharbouche L, Gil García MD, Lozano A, Hamaizi H, Martínez Galera M. Determination of personal care products in water using UHPLC-MS after solid phase extraction with mesoporous silica-based MCM-41 functionalized with cyanopropyl groups. J. Sep. Sci. 2020;43:2142–2153. doi: 10.1002/jssc.201901148

10. Zborníková E, Knejzlík Z, Hauryliuk V, Krásný L, Rejman D. Analysis of nucleotide pools in bacteria using HPLC-MS in HILIC mode. *Talanta*. 2019; 205:120161. doi: 10.1016/j.talanta.2019.120161
11. Xiao, J. F.; Zhou, B.; Ransom, H. W., Metabolite identification and quantitation in LC-MS/MS-based metabolomics. *TrAC Trends in Analytical Chemistry* 2012, 32, 1-14.
12. Tarning, J.; Bergqvist, Y.; Day, N.P.; Bergquist, J.; Arvidsson, B.; White, N. J., Characterization of human urinary metabolites of the antimalarial piperazine. *Drug Metab. Dispos.* 2006, 34 (12), 2011–2019.
13. Aziz, M. Y.; Hoffmann, K.-J.; Ashton, M., LC–MS/MS quantitation of antimalarial drug piperazine and metabolites in human plasma. *Journal of Chromatography B* 2017, 1063, 253–258.
14. Jansen, R.S.; Rosing, H.; Schellens, J.H.; Beijnen, J.H. Mass spectrometry in the quantitative analysis of therapeutic intracellular nucleotide analogs. *Mass Spectrom. Rev.* 2011, 30 (2) 321–343.
15. Gautam, N.; Lin, Z.; Banoub, M. G.; Smith, N. A.; Maayah, A. Simultaneous quantification of intracellular lamivudine and abacavir triphosphate metabolites by LC–MS/MS *Journal of Pharmaceutical and Biomedical Analysis* 2018, 153, 248-259.
16. Schuster O, Zvi A, Rosen O, et al. Specific and rapid SARS-CoV-2 identification based on LC-MS/MS analysis. *ACS Omega* 2021;6 (5):3525–3534.
17. Pavia DL, Lampman GM, Kriz GS, Vyvyan JR *Introduction to Spectroscopy*, 5th (edn), pp.120-122.
18. Steel C, Michael Henchman (1998) *Understanding the Quadrupole Mass Filter through Computer Simulation*. *J Chem Educ* 75(8): 1049-1054.
19. Guilhaus M (1995) Special Feature: Tutorial, Principles and Instrumentation in Time-of-flight Mass Spectrometry, Physical and Instrumental Concepts. *Journal Of Mass Spectrometry* 30(11): 1519-1532.
20. Raymond EM, John FT (1995) *Practical Aspects of Ion Trap Mass Spectrometry Volume-III, Chemical, Environmental and Biomedical Applications*. By CRC Press, USA, Inc p. 4-19.
21. Raymond EM, John FJ Todd (1995) Special Feature: Tutorial an Introduction to Quadrupole Ion Trap Mass Spectrometry. *Journal Of Mass Spectrometry* 32: 351-369.
22. Barnes IVJH, Hieftje GM (2004) Review Recent advances in detectorarray technology for mass spectrometry. *International Journal of Mass Spectrometry* 238(1): 33-46.
23. Kang JS (2012) Principles and Applications of LC-MS/Ms for the Quantitative Bioanalysis of Analytes in Various Biological Samples 441- 492.
24. Settle FA (2004) *Handbook of Instrumental techniques for Analytical Chemistry*. First Indian Reprint 569-660.
25. Perrenoud L, Saugy M, Saudan C (2009) Short communication Detection in urine of 4-methyl-2-hexanamine, a doping agent. *Journal of Chromatography* 877(9): 3767-3770.
26. Sargel L, Wu-Pongs, Yu ABC (2005) *Applied Biopharmaceutics and Pharmacokinetics*. 5th (edn) Mc Graw-Hill, New York, USA, p. 5-9.