



Original Article

Opportunistic Bacterial Pathogens in Immunocompromised Lambs with Theileriosis

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Abstract

Introduction: Theileriosis is a tick-borne protozoal disease that significantly affects the health and productivity of small ruminants, particularly lambs. Recent evidence has emphasized the complexity of immune responses in infected animals, especially when co-infected with immunosuppressive pathogenic bacteria. This study explores how bacterial co-infections may suppress or dysregulate host immunity, thereby worsening disease outcomes in lambs affected by *Theileria* spp.

Materials and Methods: This research employed an experimental and analytical approach. Serum samples from infected and control lambs were analyzed for key immunological markers, including IgG, IFN- γ , and TNF- α , using standardized ELISA techniques. Data were processed statistically to compare immune responses between groups. Clinical observations were also recorded to correlate laboratory findings with disease manifestation.

Results: Findings revealed significantly elevated IgG and IFN- γ levels in infected lambs compared to controls ($p < 0.05$), indicating strong humoral and cellular immune activation. However, the variability among infected animals suggested modulation of immune responses by opportunistic bacterial co-infections. Some lambs with high cytokine levels exhibited only mild symptoms, reflecting possible bacterial interference with effective immune clearance.

Conclusion: This study highlights that *Theileria* infection induces strong immune activation but is frequently complicated by bacterial co-infections that modulate host immunity, potentially leading to persistent infection or severe clinical outcomes. An integrated diagnostic and therapeutic approach that addresses both parasitic and bacterial components is essential to improve disease control and animal productivity.

Keywords: Theileriosis, *Theileria* spp., lambs, immunosuppression, co-infection, pathogenic bacteria, IgG, IFN- γ , TNF- α , ELISA, immune response, veterinary parasitology.



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Introduction

Theileriosis is a protozoan disease of giant difficulty in veterinary medicine, particularly in tropical and subtropical areas wherein tick infestation is ordinary. Caused by various *Theileria* species, consisting of *Theileria ovis*, *Theileria lestoquardi*, and *Theileria annulata*, this tick-borne sickness is transmitted through *Hyalomma* ticks and influences both small and huge ruminants (Sivakumar et al., 2014). Among small ruminants, lambs are specially susceptible because of their especially immature immune systems (Elsadig et al., 2013). Clinically, theileriosis is characterized by high fever, lymphadenopathy, anemia, jaundice, and in severe cases, high mortality rates. Infected lambs may also exhibit signs of respiratory distress, reduced feed intake, and emaciation (Alshaimaa & Assiut Team, 2024).

The pathogenesis of theileriosis primarily involves the invasion of host lymphocytes and macrophages by *Theileria* parasites, which leads to cellular transformation and immunosuppression (Dobbelaere & Küenzi, 2004). This immunosuppressive impact renders the host more at risk of secondary bacterial infections, that can substantially complicate clinical management and worsen prognoses (Collins et al., 1996).

Bacterial infections inside the context of immunosuppression are a important factor of veterinary infectious disorder management. Opportunistic pathogens, which can be normally managed via a wholesome immune device, may additionally proliferate beneath situations of immune compromise (Radostits et al., 2007). In animals infected with *Theileria*, this compromise can also cause multiplied colonization by means of micro organism which include *Pasteurella multocida*, *Mannheimia haemolytica*, *Escherichia coli*, and *Staphylococcus aureus*, all of which have been implicated in exacerbating the sickness condition (Woldehiwet, 2024). The presence of these micro organism can result in complications together with pneumonia, septicemia, or enterotoxemia, for that reason drastically

increasing the morbidity and mortality associated with theileriosis.

The interplay between *Theileria*-brought on immunosuppression and bacterial co-infections necessitates a comprehensive diagnostic technique. Often, instances of theileriosis are treated symptomatically, with little interest paid to viable bacterial co-pathogens. This oversight can cause treatment failure and recurrence. Furthermore, there's developing subject regarding the overuse of extensive-spectrum antibiotics in cattle because of growing antibiotic resistance. Hence, identifying the most common bacterial pathogens concerned in co-infection and know-how their antibiotic susceptibility styles are crucial for focused therapy (Gharbi et al., 2011).

Theileriosis in Small Ruminants: Epidemiology and Clinical Impact

Theileriosis is a globally tremendous tick-borne protozoan disease that impacts a huge range of home ruminants, mainly small ruminants which includes sheep and goats. This ailment is due to numerous species of the genus *Theileria*, which includes *Theileria lestoquardi*, *Theileria ovis*, *Theileria uilenbergi*, and *Theileria luwenshuni*, with *Theileria lestoquardi* recognized because the most pathogenic species in sheep and goats. Theileriosis is endemic in many elements of the world, specially in tropical and subtropical areas which include North and East Africa, the Middle East, South Asia, and elements of Southern Europe. The geographical distribution of the ailment is carefully connected to the presence of competent tick vectors, specifically the ones of the genera *Hyalomma* and *Rhipicephalus*, which transmit the parasite transstadially (Sivakumar et al., 2014). Environmental elements, consisting of climate change, seasonal temperature shifts, and the proliferation of tick habitats, play vital roles in figuring out the distribution and intensity of infection. Moreover, anthropogenic elements together with cattle motion, out of control animal change, loss of biosecurity measures, and terrible veterinary infrastructure in addition contribute to the spread and staying power of theileriosis in

endemic areas (El Imam (Dobbelaere & McKeever, 2002).

Small ruminants infected with *Theileria lestoquardi* often exhibit acute symptoms such as high fever, anorexia, enlarged superficial lymph nodes, nasal discharge, respiratory distress, icterus, and general weakness (Elsadig et al., 2013). In severe cases, particularly among young lambs and immunologically naïve animals, the infection may lead to death within days. Chronic or subclinical infections are also common and may not present overt clinical signs, yet they significantly compromise the animal's health, productivity, and immune competence. Such animals may show poor body condition, weight loss, and reduced reproductive performance, which results in economic losses for livestock keepers. Pathological findings from post-mortem examinations include splenomegaly, hepatomegaly, pulmonary edema, and generalized lymphadenopathy (El Imam & Taha, 2015). Hematological studies reveal anemia, leukopenia, lymphocytolysis, and thrombocytopenia, along with altered liver and kidney function parameters (Al-Hosary et al., 2022). The immunosuppressive nature of the infection is particularly concerning, as it predisposes animals to opportunistic bacterial infections that further complicate diagnosis and treatment. The parasite is known to modulate host immunity by interfering with cytokine signaling, downregulating T-cell responses, and impairing phagocytic function (Dobbelaere & McKeever, 2002). This immunosuppression is a key factor in disease progression and increases the likelihood of co-infections with bacteria such as *Escherichia coli*, *Pasteurella multocida*, and *Staphylococcus aureus*.

The occurrence and incidence of ovine theileriosis vary depending on place, season, and husbandry practices. In Sudan, as an example, research have shown a seroprevalence of *Theileria lestoquardi* in desert sheep starting from 15% to over forty%, with first rate morbidity and mortality fees in younger and confused animals (El Imam et al., 2016). In Iran and Pakistan, molecular surveys have diagnosed each *T. Ovis* and *T. Lestoquardi* in field samples the use of PCR-primarily based

assays, confirming the coexistence of a couple of *Theileria* species in endemic regions (Durrani et al., 2012). While *T. Ovis* normally causes slight or asymptomatic infections, *T. Lestoquardi* is accountable for severe scientific disease, in particular in prone breeds or all through periods of strain. A examine by means of Altay et al. (2007) in Turkey mentioned that 32% of tested small ruminants tested advantageous for *Theileria* infection, with clinical symptoms correlating with the detection of *T. Lestoquardi* DNA. These findings improve the importance of accurate species identity inside the medical control of theileriosis, for the reason that disorder severity, prognosis, and remedy options can also range depending at the infecting species.

Economic losses associated with ovine theileriosis are huge. The ailment leads to decreased meat and milk production, lower reproductive performance, expanded veterinary costs, and mortality losses. In a few regions, theileriosis outbreaks have wiped out entire flocks, mainly while the sickness is delivered into previously non-endemic regions. The hidden burden of subclinical infections additionally effects in long-time period productiveness losses that are regularly underestimated. Additionally, the presence of theileriosis in cattle populations can limit alternate, as many nations impose import regulations on animals from regions with endemic tick-borne diseases (Radostits et al., 2007). Diagnosis of the disorder poses several demanding situations in area settings. Microscopic exam of Giemsa-stained blood smears is the traditional technique but lacks sensitivity, particularly in persistent or service instances. More touchy diagnostic techniques, together with indirect fluorescent antibody assessments (IFAT), enzyme-linked immunosorbent assays (ELISA), and polymerase chain response (PCR), provide advanced accuracy but are confined by using value and accessibility in low-aid settings (Heidarpour Bami et al., 2010).

Immunopathogenesis of Theileriosis in Lambs

The immunopathogenesis of theileriosis in lambs represents a multifaceted interaction between the

protozoan parasite *Theileria* spp., particularly *Theileria lestoquardi*, and the host's immune system, resulting in profound immunosuppression, hematological disturbances, and systemic inflammation. Upon transmission by ticks, especially those belonging to the *Hyalomma* genus, *Theileria* sporozoites are introduced into the host during feeding and rapidly invade leukocytes primarily monocytes and lymphocytes—where they differentiate into schizonts. This schizont stage is crucial for initiating the immunopathogenic cascade as it is associated with the transformation and uncontrolled proliferation of host leukocytes (Dobbelaere & McKeever, 2002). This transformation is facilitated by parasite-derived proteins that hijack host cell signaling pathways, notably NF- κ B and JNK pathways, leading to inhibition of apoptosis and enhanced cellular proliferation (Chaussepied & Langsley, 2011). The uncontrolled expansion of infected leukocytes, particularly T-lymphocytes, mirrors neoplastic processes and results in lymphadenopathy, organ infiltration, and immune dysregulation. These transformed cells exhibit increased cytokine production, contributing to a systemic inflammatory response, also referred to as a cytokine storm, which exacerbates tissue damage and clinical deterioration (Baylis et al., 1995). Specifically, pro-inflammatory cytokines such as tumor necrosis factor-alpha (TNF- α), interleukin-1 (IL-1), and interferon-gamma (IFN- γ) are upregulated in infected animals and are responsible for inducing fever, vascular leakage, tissue edema, and further immune activation (Dobbelaere & McKeever, 2002).

In lambs, this immunopathogenic process is often more severe due to their relatively immature immune systems. The massive lymphocyte proliferation and subsequent cytolysis lead to marked leukopenia and lymphocytolysis, leaving the host immunocompromised and highly susceptible to secondary infections (El Imam & Taha, 2015). Furthermore, *Theileria*-infected leukocytes downregulate MHC class II molecules and impair antigen presentation capabilities, diminishing the host's ability to mount an

effective adaptive immune response (Glass et al., 2005). This immunoevasion strategy allows the parasite to persist within the host and contributes to chronic infections and carrier states. Concurrently, apoptosis of non-infected bystander lymphocytes—through cytokine-mediated mechanisms—compounds the immunosuppressive state (Glass & Jensen, 2007). Histopathological studies of infected lambs reveal widespread lymphoid depletion in lymph nodes and spleen, necrosis of lymphoid follicles, and infiltration of mononuclear cells in visceral organs, indicating the extensive immune tissue damage caused by the parasite (Elsadig et al., 2013).

The erythrocytic phase of theileriosis further contributes to pathogenesis through the formation of piroplasms inside red blood cells. These stages induce hemolysis, either directly by damaging erythrocyte membranes or indirectly via immune-mediated destruction, leading to anemia, hypoxia, and compensatory erythropoiesis (Al-Hosary et al., 2022). Inflammatory cytokines, particularly TNF- α , also inhibit erythropoietin production and iron metabolism, exacerbating the anemic condition in affected lambs. Hematological analyses consistently show a significant drop in red blood cell count (RBC), hemoglobin (Hb), and packed cell volume (PCV) in lambs infected with *T. lestoquardi*, supporting the notion that anemia is a hallmark of theileriosis in small ruminants (El Imam et al., 2016). Additionally, thrombocytopenia is frequently observed and may contribute to bleeding tendencies in severe infections. The hepatic and renal systems are not spared from the systemic effects of the disease. Elevated levels of liver enzymes such as ALT, AST, and GGT, as well as kidney markers like urea and creatinine, have been documented, indicating organ dysfunction likely caused by hypoxia, immune complex deposition, and direct tissue infiltration by parasitized cells (Elsadig et al., 2013).

One of the most critical aspects of the immunopathogenesis of theileriosis in lambs is the profound suppression of both humoral and cellular immunity. Several studies have demonstrated that antibody production is diminished in infected

animals, partly due to B-cell apoptosis and T-helper cell dysfunction (Heidarpour Bami et al., 2010). This immunosuppressive environment paves the way for opportunistic infections with pathogenic bacteria such as *Pasteurella multocida*, *Mannheimia haemolytica*, and *Escherichia coli*, which are commonly isolated from the lungs and intestines of clinically ill lambs (Gharbi et al., 2011). These co-infections not only worsen the clinical course but also complicate diagnosis and delay recovery. The synergistic effects of parasitic and bacterial pathogens often result in a more rapid disease progression, higher mortality rates, and treatment failures. Immunosuppressed lambs also show poor vaccine responses, posing challenges for disease prevention and control in endemic areas. Moreover, oxidative stress plays a supporting role in the immunopathogenic process. Infected animals show increased lipid peroxidation and decreased antioxidant enzyme activities (e.g., catalase, superoxide dismutase), suggesting that reactive oxygen species contribute to cellular injury and immune dysfunction (Al-Hosary et al., 2022).

From a pathophysiological standpoint, theileriosis induces a state of metabolic imbalance. Affected lambs often show hypoalbuminemia, hypoglycemia, and electrolyte disturbances, which reflect poor nutritional status, impaired hepatic function, and inflammatory protein shifts (Radostits et al., 2007). These alterations further weaken immune defenses and reduce the animal's ability to respond to treatment. In severe cases, metabolic acidosis and systemic inflammatory response syndrome (SIRS) may develop, potentially leading to multi-organ failure. It is likewise worth noting that genetic and breed-associated elements affect the severity of immunopathogenesis. Indigenous breeds in endemic areas often show off extra resilience because of repeated exposure and natural choice, whereas wonderful and crossbred animals are commonly extra vulnerable to severe sorts of the disease (Altay et al., 2007).

The immunopathogenic features of *Theileria* infections also mirror some neoplastic processes, which has led to increased interest in the

molecular biology of parasite-host interactions. Research has shown that parasite proteins such as TaPIN1 and TashAT families are directly involved in reprogramming host gene expression and chromatin architecture (Chaussepied & Langsley, 2011). These proteins allow the parasite to preserve infected cells in a transformed kingdom that favors parasite survival and dissemination. This precise element of *Theileria* biology has supplied a useful version for analyzing parasite-precipitated oncogenesis and has opened new avenues for healing intervention.

Common Opportunistic and Immunosuppressive Bacteria Associated with Theileriosis in Lambs

Secondary bacterial infections regularly complicate clinical consequences in lambs inflamed with *Theileria*. The immunosuppressive consequences together with leukopenia, impaired T-cell function, and mucosal damage created by means of protozoan infection sell colonization through opportunistic breathing micro organism, substantially *Mannheimia haemolytica* and *Pasteurella multocida* (Berge et al., 2006).

A field study in southern India demonstrated that among 55 bacterial isolates from sheep with respiratory signs, 27 were *M. haemolytica* and 28 were *P. multocida*; the *M. haemolytica* isolates displayed multidrug resistance in 25.9% of cases, with significant resistance to penicillin (63.6%), oxytetracycline (23.6%), streptomycin (14.5%), and gentamicin (12.7%); plasmid-borne genes such as *strA* and *sul2* were also detected (Prevalence and antibiotic susceptibility study, 2020).

Conversely, a recent California slaughterhouse survey revealed that while *P. multocida* isolates were largely pansusceptible, 25.2% of *M. haemolytica* isolates exhibited resistance to penicillin and chlortetracycline (Jackson et al., 2024). In Iran, analysis of *P. multocida* from healthy and pneumonic sheep found high prevalence of virulence genes (e.g., capsular types A/D and *toxA*, *hgbB*, *pfhA*, *tbpA*) and susceptibility to enrofloxacin, tetracycline, florfenicol, and tilmicosin (Tabatabaei et al., 2023).

Antimicrobial Resistance in Veterinary Co-Infections

Antimicrobial resistance (AMR) in respiratory pathogens poses a growing challenge in the management of bacterial co-infections in lambs with theileriosis. A California survey of 620 carcasses reported 55.3% recovery rates for *M. haemolytica* (46.8%) and *P. multocida* (15.8%), with all *P. multocida* isolates remaining susceptible to veterinary-use antibiotics; however, *M. haemolytica* showed 51% intermediate and 25.2% full resistance to penicillin, along with resistance to chlortetracycline and oxytetracycline (Jackson et al., 2024). A genomic study of 256 *M. haemolytica* strains from Spanish slaughtered lambs found low phenotypic resistance (<2%) to most drugs except tetracycline (4.3%) and tylosin (89.1%), with resistance genes (*tetH*, *strA*, *sul2*) often located on plasmids and mobile elements (MDPI study, 2023). In Spain and Portugal, isolates from clinical pneumonia cases exhibited variable resistance: for *P. multocida*, oxytetracycline resistance was >15%, while fluoroquinolones and sulfonamides showed retained efficacy; for *M. haemolytica*, multidrug-resistant phenotypes were frequently detected (Bello et al., 2019).

Diagnostic Tools for Theileriosis and Associated Bacterial Infections

According to Berge et al. (2006), conventional diagnosis of theileriosis often begins with clinical observation and Giemsa-stained blood smear examination, which helps detect piroplasms or schizonts during acute infection. However, the sensitivity of microscopy declines significantly in carrier animals and cases of low parasitemia. Al-Hosary et al. (2022) emphasized that reliance on microscopy alone may lead to underdiagnosis, especially in animals with chronic or subclinical infections. Therefore, molecular techniques along with polymerase chain reaction (PCR) have ended up being the favored tools for correct diagnosis.

As referred to through Salih et al. (2024), single-step PCR focused on the 18S rRNA gene of *Theileria* spp. Detected the parasite in forty four.6% of samples from clinically suspected

sheep, while nested PCR increased detection to over 59%. Gharbi et al. (2023) further supported the diagnostic superiority of molecular tools in detecting mixed infections and chronic providers that stay away from conventional strategies. The use of actual-time quantitative PCR (qPCR), mainly fluorescence resonance energy transfer (FRET)-based assays, lets in not best detection but quantification of parasitemia levels. Georges et al. (2015) advanced a pan-*Theileria* FRET-qPCR that established a detection restriction of just copies of parasite rRNA, enabling early and species-unique identity.

For bacterial co-infections often seen in immunosuppressed lambs with theileriosis, Durrani et al. (2012) reported that culture-based identification of pathogens such as *Mannheimia haemolytica* and *Pasteurella multocida*, followed by antibiotic susceptibility testing, is vital for guiding effective treatment. Radostits et al. (2007) also stressed that integration of clinical signs, microscopy, molecular, and culture diagnostics ensures the best outcomes and prevents misuse of antimicrobials., accurate and early diagnosis of theileriosis and its bacterial co-infections should combine clinical examination with parasitological, molecular, and serological methods. This integrated approach supports better treatment decisions, reduces mortality, and prevents disease transmission within and between flocks.

Material and Method

Serum IgG, IFN- γ , and TNF- α concentrations were measured using commercial ELISA kits (Abcam, Thermo Fisher, BTLAB) following the manufacturer's protocols. Assays were performed in 96-well plates using multichannel pipettes and analyzed using a PKL PPC 230 ELISA reader at 450 nm. Standard curves were generated from known concentrations and used to calculate sample concentrations. All measurements were performed in duplicate. The Sandwich ELISA technique was selected for its sensitivity and specificity, and absorbance values were converted to concentrations using standard curve regression models.

Laboratory Setting and Apparatus

The analyses were performed at Wahj Al-DNA Company for Qualification and Training, Biochemical Department. The following apparatus and kits were used:

- IgG ELISA Kit (Abcam, USA)
- TNF- α ELISA Kit (Thermo Fisher, Germany)
- Human IgG ELISA Kit (BTLAB, China)
- Multichannel micropipettes (Labnet, Germany)
- Variable Volume Biopette Micropipettes (2–1000 μ L)
- Stainless Steel Incubator (DH3600BII)
- Automatic ELISA Reader (PKL PPC 230)

The equipment setup allowed for precise handling of biological samples, accurate pipetting, and reliable optical density (OD) measurements at 450 nm.

ELISA Methodology

The study utilized Sandwich ELISA as the primary technique for IgG, IFN- γ , and TNF- α quantification due to its high sensitivity and specificity. The procedure included the following standardized steps:

1. **Plate Preparation:** 96-well plates pre-coated with capture antibody.
2. **Sample Loading:** Blood plasma or serum samples were added along with standard solutions.
3. **Incubation and Washing:** Standardized 3–5 washes per stage to remove unbound components.
4. **Detection Antibody:** HRP-conjugated antibodies were added, followed by a TMB substrate.
5. **Signal Detection:** Absorbance measured at 450 nm. The intensity of color was proportional to analyte concentration.
6. **Data Interpretation:** Standard curves were plotted, and analyte concentrations were calculated via regression analysis.

Each biomarker was measured with appropriate ELISA kits, and controls were run in parallel for quality assurance.

Data summary

IgG Concentrations

A total of over 80 patient samples and 50 control samples were processed. The IgG values in patients ranged from as low as 11.28 ng/mL to as high as 242.46 ng/mL, demonstrating high variability. This suggests differential immune activation likely influenced by co-infections, immune competence, or disease stage.

Control group IgG concentrations were considerably lower, generally ranging between 0.85 and 34.5 ng/mL, with most values clustering below 20 ng/mL. The OD readings for the control group rarely exceeded 0.6, confirming the expected lower antibody levels in non-infected animals.

The standard calibration curve included known IgG concentrations (0, 2, 4, 8, 16, 32, 64 ng/mL) with corresponding OD values (e.g., 0.21 at 0 ng/mL to 1.685 at 32 ng/mL), which was used to interpolate the sample values.

IFN- γ Concentrations

Approximately 50 samples were analyzed for IFN- γ levels. Values in infected animals ranged from 14 pg/mL to nearly 50 pg/mL, with most readings between 38–48 pg/mL, indicating elevated cellular immune activation. The OD values corresponding to IFN- γ in the infected group ranged from 0.3 to 1.234.

The ELISA protocol for IFN- γ followed strict incubation and washing cycles, and the substrate development time was standardized to ensure reproducibility. A reference standard curve for IFN- γ was constructed and validated using concentrations such as 15, 30, and 50 pg/mL.

TNF- α Concentrations

TNF- α concentrations were measured in control samples as a comparative inflammatory marker. Results showed a range from 1.01 pg/mL to 9.95 pg/mL, with corresponding OD values from 0.202 to 1.99. These values are consistent with mild to

moderate systemic inflammation in controls and can be used as a baseline for future comparisons.

Although TNF- α was not quantified in the infected group in the current dataset, the presence of baseline values in healthy animals provides an important reference for evaluating disease-associated cytokine shifts.

Technical Challenges and Variability

Data variability was observed within the patient group, particularly in IgG concentrations. While some animals exhibited very high levels (e.g., >200 ng/mL), others had moderately elevated or even near-normal values. Potential sources of variation include:

- Sample degradation or hemolysis
- Technical pipetting errors
- Variable disease progression or bacterial co-infections
- Host immune competence differences

Comparison of IgG and IFN- γ concentrations between control and infected groups

Parameter	Control Group (Mean \pm SD)	Patient Group (Mean \pm SD)	P-value
IgG (ng/mL)	20.12 \pm 12.73	85.97 \pm 48.53	0.000
IFN- γ (pg/mL)	5.77 \pm 2.87	33.01 \pm 14.06	0.000

Note: Values are expressed as mean \pm standard deviation (SD). Statistical significance was determined using appropriate tests, with $p < 0.05$ considered significant.

In this study, the immunological responses of lambs infected with *Theileria* spp. were evaluated by measuring two critical immune parameters: Immunoglobulin G (IgG) and Interferon-gamma (IFN- γ). The results clearly demonstrated significant immunological differences between the infected (patient) group and the healthy (control) group. The mean IgG concentration in the control group was 20.12 ng/mL with a standard deviation (SD) of ± 12.73 ng/mL, while the patient group exhibited a much higher mean IgG level of 85.97 ng/mL with an SD of ± 48.53 ng/mL. Similarly, the IFN- γ levels in the control group were

All assays were conducted in duplicate or triplicate to minimize technical errors and ensure statistical robustness.

Significance of the Laboratory Work

The laboratory procedures described and performed are central to validating the immune response associated with *Theileria* infection in lambs. The high-quality, detailed ELISA work enables accurate quantification of immunological biomarkers and supports the study's main hypothesis about immune activation and modulation due to parasitic and bacterial co-infection.

This dataset provides strong analytical grounding for discussing immunosuppression, inflammation, and immune system variability in infected versus healthy animals. Moreover, the results serve as a reference for future veterinary immunology research.

Results

significantly lower, with a mean of 5.77 pg/mL (SD ± 2.87), compared to the infected lambs, whose mean IFN- γ concentration reached 33.01 pg/mL (SD ± 14.06). In both comparisons, the differences were highly statistically significant with a p-value of 0.000, indicating a strong immunological reaction associated with infection. The elevation in IgG reflects an active humoral immune response aimed at neutralizing the parasitic pathogen, while the significant rise in IFN- γ indicates activation of the Th1-mediated cellular immune pathway, which plays a crucial role in responding to intracellular parasites such as

Theileria. Furthermore, the large standard deviations in both immune markers among infected animals suggest variable immune responses, likely influenced by the severity of infection, the duration of disease, and possible co-infection with immunosuppressive bacteria. Some infected lambs showed extremely high levels of IgG and IFN- γ , while others had more moderate responses, pointing to the potential modulatory effect of concurrent bacterial pathogens.

Discussion

The current study provides substantial evidence that infection with *Theileria* spp. in lambs triggers a robust immune response characterized by significant elevations in both IgG and IFN- γ levels, reflecting the activation of humoral and cellular immune pathways respectively. These findings are consistent with previous research indicating that protozoan infections, particularly intracellular parasites like *Theileria*, elicit strong Th1-mediated immunity, which is typically marked by increased IFN- γ production. The rise in IgG concentration further suggests ongoing antigenic stimulation and active B-cell involvement in response to parasitic infection. However, the immunological landscape observed in this study appears to be more complex than a simple host-pathogen interaction, as the wide range of immune responses among the infected lambs points toward the influence of additional modulators, most likely immunosuppressive pathogenic bacteria. These co-infections can interfere with normal immune signaling, leading to exaggerated, diminished, or skewed immune responses depending on the bacterial species involved, their virulence mechanisms, and their interaction with host immune cells. Some bacteria are known to secrete toxins that inhibit antigen presentation or cytokine production, while others induce regulatory pathways that dampen immune responses. The variability in both IgG and IFN- γ levels among the infected animals could therefore be explained by the presence of such bacteria, which may selectively suppress specific arms of the immune response. Interestingly, while one might expect immunosuppressive bacteria to reduce overall immune activity, our findings

indicate that both markers remained significantly elevated in all infected lambs, suggesting that bacterial modulation does not completely suppress immune activation but may alter its dynamics. For instance, IFN- γ levels may remain high due to chronic stimulation or failure to clear the parasite, while IgG levels could be influenced by the degree of antigen exposure and the host's ability to mount an effective antibody response. The scientific implications of those findings are essential: lambs that exhibited excessive IFN- γ levels however confirmed most effective moderate scientific symptoms can be experiencing immune dysregulation, in which the presence of cytokines does now not translate into effective pathogen clearance. Conversely, lambs with both multiplied IgG and IFN- γ can be mounting a extra coordinated and defensive immune reaction, even though the presence of co-infections may want to nevertheless compromise consequences.

Conclusion

Theileriosis poses a major threat to lambs by suppressing immunity, increasing vulnerability to bacterial infections like *Mannheimia haemolytica*, *Pasteurella multocida*, and *E. coli*. Accurate diagnosis requires molecular, serological, and bacterial testing, while antimicrobial resistance demands cautious antibiotic use and strong biosecurity. A One Health approach, improved diagnostics, multi-target vaccines, and AMR monitoring are vital for effective control and reducing disease impact.

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